Intended Use
For the quantitative determination of Albumin in serum using the Pointe c2000 and Mindray BS-200 analyzers.

Method History
Determination of serum albumin is usually made using an ultra centrifugation, salt fractionation, electrophoretic or dye binding method. Dye binding procedures are the simplest to perform, and lend themselves to high volume testing and automation. They are also the procedures most widely used in combination with total protein determinations to yield an A/G ratio. In 1953, the use of methyl orange for direct determination was described. This method suffered from non-specific binding characteristics. The use of a HABA dye was introduced in 1954. This method was specific for albumin but displayed poor sensitivity, poor correlation with electrophoresis methods and significant interference from bilirubin, lipids, salicylates, penicillin and sulfonamides. 

A bromocresol green (BCG) dye-binding procedure was first proposed in 1964. This procedure exhibited greater sensitivity and much lower susceptibility to interfering substances. The original method has been optimized to improve correlation with electrophoretic methods. The present procedure follows a modification of the original BCG dye-binding procedure.

Several publications of the late 1970's reported that abnormal proteins will bind with BCG after the first minute. The present procedures include a reduced measuring time to eliminate abnormal globulin interference and offers linearity to 8.0 g/dl.

Principle
Albumin is bound by the BCG dye to procedure an increase in the blue-green color measured at 630 nm. The color increase is proportional to the concentration of albumin present.

Reagents
Bromocresol Green (BCG) 0.15 g/L, Buffer, pH 4.66±0.1, surfactant, non-reactive ingredients and stabilizers.

Reagent Preparation
Reagent is in a “ready to use” state.

Reagent Storage
Store the reagent at room temperature (15-30°C). The reagent is stable until the expiration date appearing on the label when stored as directed.

Reagent Deterioration
The reagent should be clear, yellow-green solution. Turbidity or precipitation makes the reagent unsatisfactory and it should be discarded.

Precautions
1. This reagent is for in vitro diagnostic use only.
2. Avoid ingestion.
3. Avoid contact. Reagent is an acid solution. Flush with water when contact occurs.
4. Reagent contains Sodium Azide as a preservative. This may react with copper or lead plumbing to form explosive metal azides. Upon disposal, flush with large amounts of water to prevent azide build up.

Specimen Collection and Storage
1. Serum is the specimen of choice.
2. Avoid excessive hemolysis since every 100 mg/dl of hemoglobin corresponds to about 100 mg/dl of albumin.
3. Albumin in serum is reported stable for one week at room temperature (18-30°C) and approximately one month when stored in the refrigerator (2-8°C) and protected against evaporation.

Interferences
1. See Young et al15 for a list of interfering substances.
2. Ampicillin has been found to seriously interfere with BCG methods.16

Materials Provided
Albumin reagent.

Materials Required but not Provided
1. Analyzer.
3. Chemistry Calibrator, catalog number C7506-50
4. Chemistry Control, catalog number C7592-100

Test Parameters

<table>
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<th>Test Parameters</th>
<th>ALB</th>
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<tbody>
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<tr>
<td>Full Name</td>
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<td>Direction</td>
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Calibration Parameters

<table>
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<th>Rule:</th>
<th>Two-point linear</th>
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Limitations
1. The dye-binding properties of albumin, other than human, differ among species.  
2. Samples with values above 8.0 g/dl should be diluted with 0.9% saline 1:1, re-run, and results multiplied by 2. Samples with results below 0.5 g/dl should be done electrophoretically.
3. Severely lipemic sera should have a serum blank.
   A. Add 0.01 ml (10μl) sample to 1.0 ml deionized water and read absorbance against deionized water at 630 nm.
   B. Subtract the serum blank absorbance from the test absorbance and use the corrected absorbance in the calculations.

Calibration
Use an NIST-traceable serum calibrator. The procedure should be calibrated according to the instrument manufacturer’s calibration instructions. If control results are found to be out of range, the procedure should be re-calibrated.

Calculation (Example)
Abs. = Absorbance
Abs. of Unknown x Conc. of = Albumin (g/dl)
Abs. of Standard std.

Example: If the Absorbance of the Unknown = 0.200 and the Absorbance of the Standard is 0.19 and the Standard Concentration = 3.5, then:

\[
\text{Abs. of Unknown} \times \text{Conc. of Standard} = 0.200 \times 3.5 = 3.68 \text{ g/dl}
\]
\[
\text{Abs. of Standard} = 0.190
\]

Quality Control
The validity of the reaction should be monitored by use of normal and abnormal control sera with known albumin concentrations. Quality control requirements should be performed in conformance with local, state, and/or Federal regulations or accreditation requirements.

Expected Values\(^1\)
3.5 – 5.3 g/dl

It is strongly recommended that each laboratory establish its own normal range.

Performance
1. Linearity: 0.5 – 8.0 g/dl
2. Comparison: A study was performed between the Pointe c2000 / Mindray BS-200 and a similar analyzer and method, resulted in a correlation coefficient of 0.952 with a regression equation of \( y = 1.076x - 0.30 \) (n=29).
3. Precision: Precision studies were performed using the Pointe c2000 / Mindray BS-200 analyzer following a modification of the guidelines which are contained in NCCLS document EP5-T2.\(^1\)

<table>
<thead>
<tr>
<th>Within Run</th>
<th>Day to Day</th>
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<tr>
<td>Mean</td>
<td>S.D.</td>
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<tr>
<td>2.92</td>
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<tr>
<td>4.44</td>
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References